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TOXICOLOGY TECHNICAL PROCEDURES MANUAL	Effective Date: 31-March-2004		

#### 22 METHYLPHENIDATE AND RITALINIC ACID QUANTITATION AND CONFIRMATION BY LCMS

#### 22.1 Summary

22.1.1 Methylphenidate and its metabolite, ritalinic acid, are extracted from biological samples with an acetonitrile precipitation and analyzed by high performance liquid chromatography-electrospray ionization mass spectrometry (LC-ESI-MS).

# 22.2 Specimen Requirements

22.2.1 One mL blood, urine, gastric or tissue homogenate.

### 22.3 Reagents and Standards

- 22.3.1 Ammonium acetate
- 22.3.2 Methanol
- 22.3.3 Acetonitrile
- 22.3.4 Methylphenidate, 1 mg/mL
- 22.3.5 Ritalinic acid (α-phenyl-2-piperidineacetic acid), 1 mg/mL
- 22.3.6 Phenacetin, 1 mg/mL

## 22.4 Solutions, Internal Standard, Calibrators and Controls

- 22.4.1 10 mM Ammonium Acetate: Weight 0.38 g ammonium acetate. Transfer to 500 mL volumetric flask and QS to volume with dH<sub>2</sub>O
- 22.4.2 Working standard solution for methylphenidate and ritalinic acid (0.01 mg/mL)
  - 22.4.2.1 Pipet 100  $\mu$ l each of 1 mg/mL stock solutions of methylphenidate and ritalinic acid into a 10 mL volumetric flask and QS to volume with dH<sub>2</sub>O
- 22.4.3 Quality Control (QC) standard solution of methylphenidate and ritalinic acid (0.01 mg/mL)
  - 22.4.3.1 Pipet 100 μl each of separate 1 mg/mL stock solutions of methylphenidate and ritalinic acid (different manufacturer, lot number or preparation than calibrators) into a 10 mL volumetric flask and QS to volume with dH<sub>2</sub>O
- 22.4.4 Internal standard working solution
  - 22.4.4.1 0.1 mg/mL phenacetin: Pipet 1 mL of 1 mg/mL phenacetin stock solution into 10 mL volumetric flask and QS to volume with  $dH_2O$
- 22.4.5 To prepare the calibration curve, pipet the following volumes of the 1 mg/mL and 0.01 mg/mL methylphenidate and ritalinic acid working standards into appropriately labeled 16 x 125 mm screw cap test tubes. Add 1 mL blank blood to obtain the final concentrations listed below.

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Concentration of standard (mg/mL)	Amount of Standard ( $\mu L$ )	Final concentration of methylphenidate and ritalinic acid (mg/L)
1 mg/mL	10	10
1 mg/mL	5	5
0.01  mg/mL	200	2
0.01 mg/mL	100	1
0.01  mg/mL	50	0.5
0.01 mg/mL	10	0.1

#### 22.4.6 Controls

- 22.4.6.1 Methylphenidate and Ritalinic Acid Control
  - 22.4.6.1.1 Pipet 100 μL of the 0.01 mg/mL methylphenidate/ritalinic acid QC solution into an appropriately labeled tube. Add 1 mL blank blood to achieve final concentration of 1 mg/L.
- 22.4.6.2 Negative control. Blood bank blood or equivalent determined not to contain methylphenidate or ritalinic acid.

#### 22.5 Apparatus

- 22.5.1 Test tubes, 16 x 125 mm, round bottom, borosilicate glass with Teflon caps
- 22.5.2 Test tubes, 16 x 114 mm, glass centrifuge, conical bottom
- 22.5.3 Centrifuge capable of 2000-3000 rpm
- 22.5.4 Nitrogen evaporator with heating block
- 22.5.5 Vortex mixer
- 22.5.6 GC autosampler vials with inserts
- 22.5.7 LC/MS: Agilent Model 1100 LC-MSD
  - 22.5.7.1 LCMS Instrument Conditions. The following instrument conditions may be modified to adjust or improve separation and sensitivity.
    - 22.5.7.1.1 Elution conditions:
      - 22.5.7.1.1.1 Column: Agilent Hypersil BDS 125 mm X 3 mm, 3 μM particle size
      - 22.5.7.1.1.2 Column thermostat: 30° C
      - 22.5.7.1.1.3 Solvent A: 10 mM ammonium acetate in dH<sub>2</sub>O
      - 22.5.7.1.1.4 Solvent B: methanol
      - 22.5.7.1.1.5 Isocratic elution, stop time: 6.00 min

Time	Solv. B	Flow
0.00	48	0.5

22.5.7.1.2 Spray Chamber

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22.5.7.1.2.1 Ionization Mode: Electrospray 22.5.7.1.2.2 Gas Temperature:350° C 22.5.7.1.2.3 Drying Gas (N<sub>2</sub>): 12.0 L/min 22.5.7.1.2.4 Nebulizer pressure: 35 psig 22.5.7.1.2.5 Vcap (Positive): 3500 V

22.5.7.1.3 Selected Ion Monitoring (quantitation ions)

22.5.7.1.3.1 Polarity: Positive 22.5.7.1.3.2 Injection volume: 1 μL

Time	Group Name	SIM	Frag-	Gain	SIM	Actual
(min)		Ion	Mentor	EMV	Resol.	Dwell
0	Ritalinic acid	84	170	1.0	Low	352
		174	170		352	
		220	170		352	
2.3	Phenacetin	110	160	1.0	Low	529
		<u>180</u>	160		529	
3.5	Methylphenidate	84	170	1.0	Low	352
		174	170		352	
		<u>234</u>	170		352	

#### 22.6 Procedure

- 22.6.1 Label clean 16 x 125 mm screw cap tubes appropriately with calibrators, controls and case sample IDs.
- 22.6.2 Prepare calibrators and controls.
- 22.6.3 Add 1 mL case specimens to the appropriately labeled tubes.
- 22.6.4 Add 50 μL 0.1 mg/mL phenacetin internal standard working solution to each tube.
- 22.6.5 Slowly, add dropwise 2 mL cold (freezer temperature) acetonitrile to each tube while vortexing. Continous vortexing, not mere mixing, is essential.
- 22.6.6 Vortex an additional 30 seconds.
- 22.6.7 Place tubes in freezer for at least 30 minutes to facilitate separation.
- 22.6.8 Centrifuge at approximately 2500 rpm for 15 minutes.
- 22.6.9 Transfer top (acetonitrile) layer to clean conical bottom tubes taking care not to transfer any lower layers.
- 22.6.10 Evaporate to dryness at approximately 50° C under nitrogen.
- 22.6.11 Reconstitute samples in 100 µL methanol. Vortex briefly. Transfer to GC microvials and inject on LCMS.

#### 22.7 Calculation

22.7.1 Drug concentrations are calculated by linear regression analysis using the ChemStation software.

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22.8	8 Quality Control and Reporting				
	22.8.1	See Toxicology Quality Guidelines			
22.9	Refere	nces			
	22.9.1	J Pearson and R Steiner, in-house development.			